



## **BLOOD CLOT DNA EXTRACTION**

### **Materials needed prior to start**

- Add 96 - 100% Ethanol to Wash Buffers as indicated on the bottle.
- Set the water bath/heat block at 56°C.
- Reconstitute Proteinase K with Proteinase K Buffer and store at 2 - 8°C.

## **PROTOCOL**

### **Blood Lysate Preparation**

1. Weigh approximately 100 mg of clot and mince the blood clot using a sterile surgical blade.
2. Transfer the clot to a sterile microcentrifuge tube.
3. To the clot, add **500 µl of Blood Lysis Buffer 1** and mix thoroughly by pipetting.
4. Centrifuge at **8000 rpm for 2 mins** at room temperature.
5. Discard the supernatant and add **500 µl of Blood Lysis Buffer 1** again.
6. Resuspend the pellet by pipette mixing.
7. Centrifuge at **8000 rpm for 2 mins** at room temperature and discard the supernatant.
8. Add **500 µl of Blood Lysis buffer 2** to the pellet and resuspend by pipette mixing.
9. Add **20 µl of Proteinase K** and resuspend by pipetting.
10. Incubate at **56°C for 30 mins**.
11. Transfer the lysate to a fresh microcentrifuge tube.

### **DNA Binding and Washing**

*Note: Vortex the Blood MagNa Mix to ensure complete dispersion of the particles.*

12. Add **350 µl of Blood MagNa Mix** and invert the tube 10 - 12 times to mix properly.
13. Incubate the samples at room temperature for 5 mins.
14. Place the tube on MagNa Stand until the solution gets cleared. Discard the supernatant.
15. To the pellet, add **500 µl of Blood Wash Buffer 1**, remove the tube from MagNa Stand and resuspend by pipette mixing (*ensure complete dispersion of the particles*).

*Note: Use 200 µl pipette for better resuspension of the pellet.*



16. Place the tube back on MagNa Stand until the solution gets cleared (*30 sec - 1 min*).
17. Discard the supernatant without removing the tube from MagNa Stand (*Make sure that the pellet is not disturbed*).
18. Add **500 µl of Blood Wash Buffer 2** & gently invert mix the tube without removing from MagNa Stand for 5 - 6 times (*surface wash only*).
19. Discard the supernatant without removing the tube from MagNa Stand.
20. Repeat steps 17 - 19.
21. Air dry the pellet without removing the tube from MagNa Stand for 10 - 15 mins.

*Note: Do not over dry the pellet.*

#### **DNA Elution**

22. To the pellet add **50 µl Elution Buffer**, remove the tube from MagNa Stand, and disperse the pellet thoroughly by pipette mixing.
23. Incubate at **56°C for 5 mins**.
24. Place the tube back on MagNa Stand until the solution gets cleared.
25. Carefully transfer the supernatant to a new microcentrifuge tube and discard the MagNa particles.

### Downstream applications

DNA obtained from this method is ready to use for any downstream application without any further precipitation or purification step.

### Troubleshooting Guide

| Observation                          | Possible Causes   | Suggested Solution   |
|--------------------------------------|---|--|
| <b>Low DNA yield or Poor Quality</b> | Incomplete Lysis  | Ensure that the blood clot is minced properly and it is not too old/dried. Check if the sample is stored properly.   |
|                                      |   | Use suggested amount of Proteinase K for the specified time.   |
|                                      |   | Ensure proper re-suspension of pellet while adding lysis buffers.  |
|                                      |   | Make sure that the Lysis incubation temperature and time is followed correctly.  |
|                                      |   | In some cases when DNA content is very high, the lysate may appear slimy. Ensure thorough pipette mixing during lysis and before adding MagNa Mix in such circumstances. |
|                                      | Incorrect reagent volumes were used                             | Use the exact volumes of reagents mentioned in the protocol.   |
|                                      | MagNa Mix was improperly handled                                | Resuspend the MagNa Mix by vortexing prior to use.   |
|                                      | MagNa Mix was disturbed or lost during binding or washing steps | Carefully remove the supernatant from the tube without removing the tube from the MagNa Stand. Make sure that MagNa particles are not disturbed.                         |
|                                      | Improper elution  | Completely resuspend the MagNa particles in elution buffer before  |

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|---|--------------------------------------|---|
|   |                                      | incubation at 56°C for elution.   |
|   | Ethanol is not added to wash buffers | Add 100% ethanol to wash buffers prior to use as mentioned on the bottles.  |
| <b>Poor performance of extracted DNA in downstream applications</b> | Ethanol carryover                    | Air dry the MagNa particles properly after washing steps, to remove the ethanol completely, but do not over dry the pellet. |
|   | Salt carryover                       | Ensure that the correct amount of ethanol added to the Wash Buffers and two wash steps are performed with Wash Buffer 2.    |